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<p>(21) International Application Number: PCT/EP98/00816</p> <p>(22) International Filing Date: 12 February 1998 (12.02.98)</p> <p>(30) Priority Data: MI97A00363 20 February 1997 (20.02.97) IT</p> <p>(71) Applicant (<i>for all designated States except US</i>): ANGELINI RICERCHE S.P.A. SOCIETA' CONSORTILE [IT/IT]; Piazzale della Stazione, I-00040 S. Palomba-Pomezia (IT).</p> <p>(72) Inventors; and</p> <p>(75) Inventors/Applicants (<i>for US only</i>): CAVALLO, Giovanni [IT/IT]; Via delle Tartane, 45, I-00122 Ostia (IT). MARCITTO, Leonardo [IT/IT]; Via S. Margherita, 2, I-63012 Cupra Marittima (IT).</p> <p>(74) Agents: MARCHI, Massimo et al.; Marchi & Partners s.r.l., Via Pirelli, 19, I-20124 Milano (IT).</p>		<p>(81) Designated States: AL, AU, BA, BB, BG, BR, CA, CN, CU, CZ, EE, GE, GW, HU, ID, IL, IS, JP, KP, KR, LC, LK, LR, LT, LV, MG, MK, MN, MX, NO, NZ, PL, RO, SG, SI, SK, SL, TR, TT, UA, US, UZ, VN, YU, ARIPO patent (GH, GM, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).</p> <p>Published <i>With international search report.</i> <i>Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i></p> <p>(54) Title: AQUEOUS PHARMACEUTICAL COMPOSITION COMPRISING AN ACTIVE INGREDIENT WHICH IS HIGHLY INSOLUBLE IN WATER</p> <p>(57) Abstract Aqueous pharmaceutical composition comprising an active ingredient which is highly insoluble in water, dispersed in liposomes and method of preparation thereof.</p>	

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"Aqueous pharmaceutical composition comprising an active ingredient which is highly insoluble in water"

This invention relates to an aqueous pharmaceutical composition comprising an active ingredient which is highly insoluble in water. In particular, it relates to a pharmaceutical composition in which the active ingredient is dispersed in liposomes.

A great deal of research is being carried out to find new liposomal preparations in the pharmaceutical field. However, many difficulties have emerged particularly in relation to active ingredients which are highly insoluble in water. In particular, those with a solubility in water $\leq 0.01\%$ (w/v).

In fact, the technique currently used to produce liposomes comprising active ingredients of low water-solubility comprises:

- a) solubilizing of the active ingredient and the preselected phospholipids in a suitable organic solvent, for example, chloroform;
- b) evaporation of this solvent at reduced pressure to give an active ingredient/phospholipid film;
- c) addition of a second organic solvent, for example, terbutylic alcohol;
- d) freezing of the solution obtained at the temperature of liquid nitrogen;
- e) lyophilisation of the frozen solution;
- f) hydration of the lyophilised solution with a buffer solution to give a suspension of multilamellar liposomes (MLV); and
- g) treatment of this suspension with ultrasound to give a suspension of smaller liposomes (SUV).

An example of this method is described by A. Sharma et al.

"Pharmaceutical Research", 2 (6), 889-896 (1994):

This technique, however, has the disadvantage of being very laborious and leads to the presence of traces of organic solvents in the liposomes.

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However, the said authors refer to having investigated various techniques for the preparation of MLV liposomes such as the hydration of the dry lipid films (hand shaking), freeze thawing and various techniques such as extrusion and treatment with ultrasound to reduce then (postprocessing) the size of the liposomes (MLV → SUV) and conclude that the method described in detail above and comprising the said phases a) to g) was demonstrated to be the most acceptable (*loc. cit.* page 890, right column, lines 51-57). However, the said authors do not indicate how the said first techniques for the preparation of MLV liposomes and the said second techniques which can reduce the size of the liposomes were combined one another.

Surprisingly, it has now been found that the freezing and thawing technique combined with extrusion allows the preparation of aqueous liposome compositions of active ingredients with a solubility in water \leq 0.01% (w/v) without the use of any organic solvent.

In this description, and the claims that follow, active ingredients with a solubility in water \leq 0.01% (w/v) are defined as "highly insoluble in water".

Therefore, the first object of this invention is an aqueous composition characterised by the fact that it comprises an active ingredient, which is highly insoluble in water, dispersed in liposomes.

The following are typical examples of active ingredients which are highly insoluble in water: ionidamine (solubility: 3×10^{-6} g/ml), melatonin ["practically insoluble", G.S. Shida et al. "J. Pineal Res.", 16, 198-201, (1994)], cyclosporin-A ["insoluble in water", monograph on cyclosporin-A in "Analytical Profiles of Drug Substances", 16, 163, (1987)] and bindarit (solubility: 1×10^{-4} g/ml).

The liposomes of the compositions according to this invention are preferably made up of a component chosen from the group comprising phosphoglycerides, glycerides, diglycerides, triglycerides, phospholipids, galactosyl and glucosyl lipids, cholesterol and its derivatives,

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sphingolipids and mixture thereof. More preferably, they are made up of phospholipids.

A typical example of the liposomal composition according to this invention comprises phosphatidylcholine, lysophosphatidylcholine, N-acyl-phosphatidylcholine, phosphatidyl ethanolamine, phosphatidylserine, sphingomyelin, non-polar lipids, triglycerides, free fatty acids, DL- α -tocopherol.

A preferred liposomal composition according to this invention comprises:

<u>Component</u>	<u>% (w/w)</u>
phosphatidylcholine	: 85 - 97
lysophosphatidylcholine	: 0 - 5
N-acyl-ethanolamine	: 0 - 4
phosphatidyl ethanolamine	: 0 - 10
triglycerides	: 0 - 4
free fatty acids	: 0 - 3
DL- α -tocopherol	: 0 - 1

A particularly preferred liposomal composition according to this invention comprises:

<u>Component</u>	<u>% (w/w)</u>
phosphatidylcholine	: 94
lysophosphatidylcholine	: 3
N-acyl-ethanolamine	: 1
phosphatidyl ethanolamine	: 0.1
triglycerides	: 1
free fatty acids	: 0.75
DL- α -tocopherol	: 0.15

Typically, the size of the liposomes according to this invention is less than 500 nm. Preferably, this is from 50-250 nm.

A second object of this invention is a method for the preparation of an aqueous pharmaceutical composition with an active ingredient which is highly insoluble in water, dispersed in liposomes, which is characterised by the fact that it comprises the following phases:

- a) dispersion of this active ingredient in lipids at a temperature of between 20 and 30°C;
- b) suspension of this dispersion in an aqueous phase;
- c) resting of this suspension at ambient temperature for a period of between 0 and 48 hours;
- d) heating to 30-75°C for 10-40 minutes;
- e) freezing at -150/-200°C;
- f) repetition of phases d) and e) at least twice and not more than 8 times;
- g) filtration through a filtering membrane with pores of diameter 500-1000 nm;
- h) extrusion through a membrane with pores of diameter 50-400 nm; and at the same time
- i) removal of any active ingredient which is not trapped.

The duration of phase c) depends on the quantity of active ingredient highly insoluble in water to be trapped in the liposomes. The person skilled in the art does not therefore encounter any difficulties since a few simple routine experiments will determine the correct time for each type of active ingredient and liposomal composition.

The aqueous phase shall preferably be made up of an aqueous solution of sodium chloride at 0.05%-0.9% (w/v).

Typically, the quantity of lipid used is between 0.01-0.4 parts by weight for each part by weight of aqueous solution. In turn, the quantity of active ingredient is typically between 0.01 and 0.3 parts by weight for each part by weight of lipid.

Typically, the disperser is a homogeniser of the Ultraturrax™ type.

Typically, the extrusion is carried out using compressed air or an inert gas, chosen from the group comprising nitrogen, helium and argon, as the extrusion gas. The preferred inert gas is helium. In the extrusion phase, the pressure shall preferably be between 500 and 5500 kPa and the temperature shall preferably be between 20 and 75°C, and even more preferably between 40 and 65°C. Typical examples of suitable extruders are those of the Lipex Biomembranes Thermobarrel type or of the Emulsiflex CC Avestin type with filters with polycarbonate Costar™ membranes with pores of between 50 and 600 nm.

Typically phase h) is repeated at least twice and not more than 8 times. Preferably 6 times.

The following examples illustrate this invention without limiting it in any way.

EXAMPLE 1

100 mg of melatonin were dispersed in 1 g of phospholipid at 30°C for 10 minutes using an Ultraturrax™ type homogeniser. Immediately afterwards, this dispersion was suspended in 10 ml of aqueous solution of sodium chloride at 0.9% (w/v) using the said homogeniser and then heated in a water bath at 55°C for 20 minutes.

The suspension obtained in this way was subject to the following cycle of cooling and heating:

- cooling in liquid nitrogen for 1 minute,
- heating to 55°C until the phospholipids are completely fluid.

This cycle was repeated 6 times.

The suspension was passed twice through a 0.6 µm filter with the Lipex Biomembrane apparatus.

Thus, a "Multilamellar Large Vesicles" (MLV) suspension was obtained which was subjected to 6 cycles of continuous extrusion using a 10 ml extruder of the Lipex Biomembranes Extruder Thermobarrel type with 0.1

µm polycarbonate Costar™ filters at 55°C, using helium, as the extrusion gas, at a pressure of between 1000 and 4800 kPa.

Operating as described above three batches of the product (LM/186, LM/188 and LM/190) were prepared.

The following tests were carried out on the batches:

- * melatonin amount in the aqueous liposomal composition (HPLC analysis);
- * liposome size;
- * quantity of melatonin trapped in the liposomes.

The following table shows the parameters measured and their significance:

Parameters	Significance
liposome size	<ul style="list-style-type: none"> – stability in the formulation time; – measurement of the "fusion" of the vesicles;
melatonin amount	<ul style="list-style-type: none"> – concentration of melatonin in the aqueous liposomal composition; – stability in the formulation time;

The data obtained are given in Table 1 which shows:

- the concentration of melatonin obtained in the aqueous liposomal formulation was, expressed as an average value for the three batches, 8.05×10^{-3} g/ml;
- the average size of the liposomes for the three batches was 93 nm;
- the quantity trapped, expressed as an average value for the three batches, was 80.5 µg/mg;
- the formulations showed no liposome aggregation phenomena.

TABLE 1

	HPLC amount (mg/ml)	average size (nm)	quantity trapped (µg/mg) *
LM/186	7.8	85	78
LM/188	8.46	97	84.6
LM/190	7.9	98	79

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(*) expressed as γ of drug per mg of phospholipids used.

The following procedure was used for the HPLC analysis:

- fixed phase: column in inverse phase PKB-100 (250 x 4.6 mm; 5 μ m Supelco);
- mobile phase: water:acetonitrile 80:20 (v/v);
- detection: UV 254 nm.

Two pieces of apparatus are used for the analysis of the average size of the liposomes:

- 1) DELSA 440 Coulter,
- 2) NICOMP Submicron particle sizer model 370.

The procedure was as follows:

- a) for the tests carried out with apparatus 1), 1 ml of liposomal suspension was diluted with 10 ml of aqueous solution of sodium chloride at 0.9% (w/v);
- b) for the tests carried out with apparatus 2), 0.5 ml of solution a) was diluted to 10 ml with aqueous solution of sodium chloride at 0.9% (w/v).

EXAMPLE 2

Proceed as described in Example 1 above, using 2 g of phospholipid and 50 mg of Ionidamine in place of 1 g of phospholipid and 100 mg of melatonin.

Thus three batches of the product (LM/195, GN/1L and GN/2L) are prepared. The data obtained are given in Table 2 which shows:

- the concentration of Ionidamine in the aqueous composition went from the initial solubility value of 3×10^{-6} g/ml to an average value for the three batches of 3.83×10^{-3} g/ml;
- the average size of the liposomes for the three batches was 79.6 nm;
- the quantity trapped, expressed as an average value for the three batches, was 19.2 γ /mg;
- the formulations showed no liposome aggregation phenomena.

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TABLE 2

batch	HPLC amount (mg/ml)	average size (nm)	quantity trapped (γ /mg)*
LM/195	3.66	103	18.3
GN/1L	3.31	53	16.5
GN/2L	4.54	76	22.7

(*) expressed as γ of drug per mg of phospholipids used.

EXAMPLE 3

Proceed as described in Example 1 above, using 2 g of phospholipid and 200 mg of melatonin in place of 1 g of phospholipid and 100 mg of melatonin.

Thus three batches of the product (GN/1M, GN/2M and GN/3M) were prepared. The data obtained are given in Table 3 which shows:

- the concentration of melatonin in the aqueous liposomal formulation, expressed as an average value for the three batches, was 13.5×10^{-3} g/ml;
- the average size of the liposomes for the three batches was 92.6 nm;
- the quantity trapped, expressed as an average value for the three batches, was 67.6 γ /mg;
- the formulations showed no liposome aggregation phenomena.

TABLE 3

batch	HPLC amount (mg/ml)	average size (nm)	quantity trapped (γ /mg)*
GN/1M	10.66	104	53.3
GN/2M	13.90	76	69.5
GN/3M	16.03	98	80.15

(*) expressed as γ of drug per mg of phospholipids used.

EXAMPLE 4

Proceed as described in Example 2 above, except that the extrusion is carried out through a polycarbonate membrane of 0.2 μm rather than 0.1 μm .

Thus three batches of the product (GN/3L, GN/4L and GN/5L) were prepared.

The data obtained are given in Table 4 which shows that, by increasing the Ionidamine from 20 mg to 50 mg, the quantity of phospholipid for 1 to 2 g and extruding with a 0.2 μm instead of a 0.1 μm membrane, a significant increase in the concentration of Ionidamine in the aqueous composition was obtained in example 2. In fact, an average value of 4.47×10^{-3} g/ml was obtained for the concentration of Ionidamine.

TABLE 4

batch	HPLC amount (mg/ml)	average size (nm)	quantity trapped (γ/mg)*
GN/3L	4.23	134	21.15
GN/4L	4.44	129	22.20
GN/5L	4.75	109	23.75

(*) expressed as γ of drug per mg of phospholipids used.

EXAMPLE 5

20 mg of cyclosporin-A were dispersed in 1 g of phospholipid at 30°C for 10 minutes using an Ultraturrax™ type homogeniser. Immediately afterwards, this dispersion was suspended in an aqueous solution of sodium chloride at 0.9% (w/v) using the said homogeniser and then heated in a water-bath at 65°C for 20 minutes.

The suspension obtained in this way was subject to the following cycle of cooling and heating:

- cooling in liquid nitrogen for 1 minute,
- heating to 65°C until the phospholipids are completely fluid.

This cycle was repeated 6 times.

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The suspension was passed twice through a 0.6 μm filter with the Lipex Biomembrane apparatus.

Thus a "Multilamellar Large Vesicles" (MLV) suspension was obtained which was subjected to 6 cycles of continuous extrusion using a 10 ml extruder of the Lipex Biomembrane Extruder Thermobarrel type with 0.1 μm polycarbonate CostarTM filters at 65°C, using helium as the extrusion gas at a pressure of between 1000 and 4800 kPa.

Thus three batches of the product (LM/416A, LM/416B and LM/416C) were prepared.

The data obtained are given in Table 5 which shows:

- the concentration of cyclosporin-A in the aqueous liposomal formulation, expressed as an average value for the three batches, was 0.96×10^{-3} g/ml;
- the average size of the liposomes for the three batches was 103 nm;
- the quantity trapped, expressed as an average value for the three batches, was 9.6 γ/mg ;
- the formulations showed no liposome aggregation phenomena.

TABLE 5

batch	HPLC amount (mg/ml)	average size (nm)	quantity trapped (γ/mg)*
LM/416A	0.96	103	9.6
LM/416B	0.94	99	9.4
LM/416C	0.98	107	9.8

(*) expressed as γ of drug per mg of phospholipids used.

EXAMPLE 6

Proceed as described in Example 1 above, using 2 g of phospholipids and 50 mg of bindarit in place of 1 g of phospholipids and 100 mg of melatonin.

Thus three batches of the product (LM/356, LM/357 and LM/358) were prepared.

The data obtained are given in Table 6 which shows:

- the concentration of bindarit in the aqueous liposomal composition went from the initial solubility value of 1×10^{-4} g/ml to an average value for the three batches of 4 mg/ml;
- the average size of the liposomes for the three batches was 108.3 nm;
- the quantity trapped, expressed as an average value for the three batches, was 20.2 $\mu\text{g}/\text{mg}$;
- the formulations showed no liposome aggregation phenomena.

TABLE 6

batch	HPLC amount (mg/ml)	average size (nm)	quantity trapped ($\mu\text{g}/\text{mg}$)*
LM/356	4.1	109.4	20.5
LM/357	4	109.7	20
LM/358	4	106	20

(*) expressed as μg of drug per mg of phospholipids used.

EXAMPLE 7

30 mg of cyclosporin-A were dispersed in 2 g of phospholipid at 30°C for 10 minutes using an Ultraturrax™ type homogeniser. Immediately afterwards, this dispersion was suspended in an aqueous solution of sodium chloride at 0.9% (w/v) using the said homogeniser and left to rest at ambient temperature for 24 hours. Then the suspension obtained was heated in a water-bath at 65°C for 20 minutes.

The suspension obtained in this way was subject to the following cycle of cooling and heating:

- cooling in liquid nitrogen for 1 minute,
- heating to 65°C until the phospholipids are completely fluid.

This cycle was repeated 6 times.

The suspension was passed twice through a 0.6 μm filter with the Lipex Biomembrane apparatus.

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Thus, a "Multilamellar Large Vesicles" (MLV) suspension was obtained which was subjected to 6 cycles of continuous extrusion using an extruder of the 10 ml Lipex Biomembrane Extruder Thermobarrel type with 0.1 μ m polycarbonate CostarTM filters at 65°C, using helium as the extrusion gas at a pressure of between 1000 and 4800 kPa.

Thus three batches of the product (LM/422a, LM/422b and LM/422c) were prepared.

The data obtained are given in Table 7 which shows:

- the concentration of cyclosporin-A in the aqueous liposomal formulation, expressed as an average value for the three batches, was 3 mg/ml;
- the average size of the liposomes for the three batches was 119.5 nm;
- the quantity trapped, expressed as an average value for the three batches, was 15 γ /mg;
- the formulations showed no liposome aggregation phenomena.

TABLE 7

batch	HPLC amount (mg/ml)	average size (nm)	quantity trapped (γ /mg)*
LM/422a	3.2	121.5	16
LM/422b	3	117.9	15
LM/422c	2.8	119	14

(*) expressed as γ of drug per mg of phospholipids used.

CLAIMS

1. An aqueous pharmaceutical composition characterised in that it comprises an active ingredient highly insoluble in water, dispersed in liposomes.
2. A composition according to claim 1, characterised in that the active ingredient highly insoluble in water is chosen from the group comprising lonidamine, melatonin, cyclosporin-A and bindarit.
3. A composition according to claim 1 or 2, characterised in that the liposomes are constituted of a component chosen from the group comprising phosphoglycerides, glycerides, diglycerides, triglycerides, phospholipids, galactosyl and glucosyl lipids, cholesterol and its derivatives, sphingolipids and mixture thereof.
4. A composition according to any of claims 1 to 3, characterised in that the liposomes are constituted by a composition comprising phosphatidylcholine, lysophosphatidylcholine, N-acyl-phosphatidylcholine, phosphatidyl ethanolamine, phosphatidylserine, sphingomyelin, non-polar lipids, triglycerides, free fatty acids, DL- α -tocopherol.
5. A composition according to any of claims 1 to 4, characterised in that the quantity of lipids is between 0.01 and 0.4 parts by weight for each part by weight of water.
6. A composition according to any of claims 1 to 5, characterised in that the quantity of the active ingredient is between 0.01 and 0.3 parts by weight for each part by weight of lipids.
7. A method for the preparation of an aqueous pharmaceutical composition with an active ingredient highly insoluble in water, dispersed in liposomes, characterised in that it comprises the following phases:
 - a) dispersion of this active ingredient in lipids at a temperature of between 20 and 30°C;

- b) suspension of this dispersion in an aqueous phase;
- c) resting of this suspension at ambient temperature for a period of between 0 and 48 hours;
- d) heating to 30-75°C for 10-40 minutes;
- e) freezing at -150/-200°C;
- f) repetition of phases d) and e) at least twice and not more than 8 times;
- g) filtration through a filtering membrane with pores of diameter 500-1000 nm;
- h) extrusion through a membrane with pores of diameter 50-400 nm; and at the same time
- i) removal of any active ingredient which is not trapped.

8. A method according to claim 7, characterised in that the aqueous phase is constituted by the aqueous solution of sodium chloride at 0.05%-0.9% (w/v).
9. A method according to claim 7 or 8, characterised in that the quantity of lipids used is between 0.01 and 0.4 parts by weight for each part by weight of water.
10. A method according to any of claims 7 to 9, characterised in that the quantity of the active ingredient used is between 0.01 and 0.3 parts by weight for each part by weight of lipids.
11. A method according to any of claims 7 to 10 above, characterised by the use in phase h) of an extrusion gas chosen from the group comprising air, nitrogen, helium and argon.
12. A method according to claim 11, characterised in that the extrusion gas has a pressure of between 500 and 5500 kPa.
13. A method according to any of claims 7 to 12 above, characterised in that phase h) is carried out at a temperature of between 20 and 75°C.

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14. A method according to claim 13, characterised in that the temperature is between 40 and 65°C.
15. A method according to any of claims 7 to 13 above, characterised in that phase h) is repeated at least twice and not more than 8 times.

INTERNATIONAL SEARCH REPORT

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A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 A61K9/127

According to International Patent Classification(IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

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C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category ³	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 96 40064 A (NEXSTAR PHARMACEUTICALS) 19 December 1996 see page 1, line 1 - page 10, line 2 ---	1-6
X	EP 0 578 620 A (I.F.L.O. S.A.S. DI GIORGIO E ALDO LAGUZZI) 12 January 1994 see claim 18 ---	1,2
X	DE 40 38 075 C (B. BRAUN MELSUNGEN AG) 19 March 1992 see page 3, line 4 - page 5, line 30 see claims 11,13 ---	1-6
A	DE 44 30 593 A (MAX-DELBRÜCK-CENTRUM FÜR MOLEKULARE MEDIZIN) 22 February 1996 see page 3, line 17 - line 53 see claim 14 ---	7-15
A	DE 44 30 593 A (MAX-DELBRÜCK-CENTRUM FÜR MOLEKULARE MEDIZIN) 22 February 1996 see page 3, line 17 - line 53 see claim 14 ---	7-14
		-/-

Further documents are listed in the continuation of box C.

Patent family members are listed in annex.

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INTERNATIONAL SEARCH REPORT

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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	EP 0 671 156 A (ROUSSEL UCLAF) 13 September 1995 see page 10; example 3 -----	7-14

INTERNATIONAL SEARCH REPORT

Information on patent family members

In' tional Application No

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